

hours, its use provides a cheap, easy way to tag flies in order to study ecologically important aspects of their biology such as feeding choices and dispersal distances *in-situ*.

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Fresh yeast media for *Drosophila* egg-collecting.

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Fertility and fecundity studies in *Drosophila* have described several egg-collecting techniques with different media as cornmeal agar or banana agar added with animal charcoal for contrast, sprayed with a fine yeast suspension (Spencer, 1937), with colored Carpenter's medium and fresh or dead yeast (Gupta, 1980) or yeast-agar with grape juice (Nichols and Pak, 1985). Egg-collecting media has been added to watch glasses (Delcour, 1969), to spoons (Gupta, 1980), or to Petri dishes with small drops of fresh yeast (Nüsslein-Volhard, 1977; Sabio *et al.*, 2010). For more contrasting of eggs some techniques include small drops of food coloring (Gupta, 1980) or vital stain (Acosta *et al.*, 2000). Any kind of egg-collecting media and vials must have these requests: i) to allow collection of large quantities of eggs; ii) to make accessible microscopic observation and incubation; iii) to assure enough amount of food for parents and larvae (Acosta *et al.*, 2000). In order to describe a method that combines the features described above, we present a simple method that allows a short preparation time, low cost, besides eggs-collecting, observation facilities and incubation efficiency. Centrifuge plastic tubes of 90 × 30 mm with removable plastic cap, and without thread, to which the conical base was cut (Acosta *et al.*, 2000) to replace with a foam plug (Figure 1) were used for egg collecting. The removable cap of the tube is completely filled with fresh yeast, previously sucrose activated and incubated in a water bath at 39°C for 5 min in three successive periods. After placing the fresh yeast in the cap with a sharp object slits are made on the surface (Figure 2). It takes 400 g of fresh yeast to prepare 80 caps with this media. Couples of males and female virgins are placed in vials for 5-6 h without food to prevent egg-laying (Delcour, 1969). About 2 h after placing the media in the removable caps, couples are placed in the vials for egg-collecting, and the tubes are closed with the foam caps. Then the tubes are placed in an incubator at 25°C and 60% relative humidity for 24 h. This method has the following advantages: i) fresh yeast promotes egg-laying and gives parents and larvae enough food; ii) yeast color contrasts with the color of the eggs, preventing exposure to dyes (Figure 3); iii) the media remains with the characteristics required for 72 h; iv) it is easy to remove the cap for complete stereo microscope observation of the media's surface; iv) caps can be replaced for continuous egg-collecting every 24 h, and caps with eggs can be incubated in new tubes.

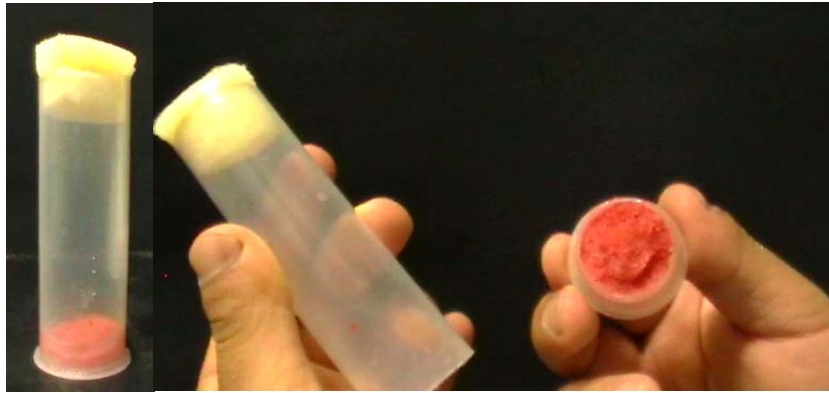


Figure 1. Centrifuge plastic tubes with removable plastic cap, and without thread, were used for egg collecting.



Figure 2. Fresh yeast, sucrose activated, is placed in the vials caps and with a sharp object slits are made on the surface.



Figure 3. The oviposition is improved by slits in the yeast media and the natural color facilitates the counting of eggs.

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